

Original Research Article

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Antibiogram of Pathogenic Urinary *E.coli* Isolates from Tertiary Care Hospital in Kerala, India

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ABSTRACT

Urinary tract infections are one of the most common causes of patient attendance in hospitals. *E.coli* remains the most commonly isolated pathogen causing Urinary tract infections. Antibiotic resistance is an emerging problem in developing countries. The way ahead is to streamline the utilisation of antibiotics and to implement locally relevant antibiotic policy. A total of 1152 urine samples were received during the study period. 327 samples grew monobacterial significant bacteriuria. Semi-quantitative cultures were performed. The isolates were tested for sensitivity to the following discs Ampicillin (10µg), Gentamicin (10µg), Tobramycin (10µg), Amikacin (30µg), Ciprofloxacin(5µg) Levofloxacin (5µg), Cotrimoxazole (1.25/23.75µg) Amox-Clavulanic Acid (20/10µg) Piperacillin-Tazobactam (100/10µg) Norfloxacin (10µg) Amikacin (30µg) Cefuroxime (30µg), Ceftriaxone (30µg), Ceftazidime (30µg), Ceftazidime- Clavulanic Acid (30/10µg), Cefotaxime (30µg) Imipenam (10µg), Tetracycline (30µg), Nitrofurantoin (300µg). The sensitivity was recorded and reported as per the CLSI 2017 guidelines. Antibiotic sensitivity profile of *E.coli* shows sensitivity to Imipenam (91%), Nitrofurantoin (97%), Amikacin (91%), Tobramycin (72%), Piperacillin-Tazobactam (77%), Gentamicin (66%) and Tetracycline (46%). *E.coli* isolated exhibited resistance to ampicillin, amoxicillin-clavulanic acid, 2nd and 3rd generation cephalosporins and fluoroquinolones. *E.coli* was sensitive to cotrimoxazole in 49% isolates. 62 (48.1%) isolates were resistant to 3 or more antibiotic groups (MDR). 17 of the 148 isolates exhibited Extended drug resistant (XDR) *E.coli* showed high degree of resistance to cephalosporins and fluoroquinolones. One of the short term measures advised is antibiotic cycling to prolong the usefulness of live saving drugs. For this every healthcare unit needs to develop their own antibiogram to have a clear picture of the antibiotic resistance pattern prevalent in the area.

Keywords

Urinary *E.coli*, Antibiotic resistance, *E.coli*, Urinary tract infection, Antibiotic sensitivity profile

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Introduction

Urinary tract infections (UTI) are the most common cause of patients attending hospital. Uncomplicated symptomatic UTI presenting with dysuria, fever and lower abdominal pain especially in female patients is a common

scenario in India. Urine sample remains the most frequent sample submitted to Microbiology laboratory for identification of offending pathogen. The main cause of urinary tract infection in India remains to be the members of Enterobacteriaceae family. This is followed by members of Pseudomonas

family, *Staphylococcus aureus* and *Enterococcus sp.* Among the members of Enterobacteriaceae *Escherichia coli* remains the most important pathogen causing uncomplicated UTI. *E. coli* infections are more common in women which may be attributed to the anatomical proximity to the gastrointestinal tract which harbours *E.coli* as a commensal (Richard Colgan and Mozella Williams, 2011; Mohammed Akram *et al.*, 2007). *Klebsiella pneumoniae* is the next most commonly isolated gram negative bacteria from Enterobacteriaceae family. Few infections are caused by *Klebsiella oxytoca*, *Proteus mirabilis*, *Proteus vulgaris* and *Citrobacter sp.* Among the gram positive bacteria *Staphylococcus aureus* and *Enterococcus* are responsible for few uncomplicated UTI's. Coagulase Negative Staphylococci (CoNS) remain a gray area in microbiological diagnosis of UTI. Being the part of skin commensals CoNS are not generally reported as an offending organism. In the recent years there have been reports of CoNS causing clinically relevant infections. CoNS infections are more common in immunocompromised population but there have been reported studies on capability of CoNS in causing infections in immunocompetent people as well. Members like *Staphylococcus hemolyticus*, *Staphylococcus lugdunensis* and *Staphylococcus saprophyticus* are notorious in causing human infections. The role *Pseudomonas aeruginosa* in causing UTI is well known. Being a hardy organism capable of growing on varied substrates including disinfectants are seen pan environmentally the isolation of *Pseudomonas aeruginosa* needs correlation with clinical condition of the patient. Nevertheless the role of *Pseudomonas aeruginosa* in causing UTI cannot be underplayed.

Antibiotic resistance has been emerging in India (Muktikesh Dash *et al.*, 2013). It has reached a stage of being one of the most

important public health problems which needs to be tackled. Since the reason for spreading antimicrobial resistance is multi-factorial, there needs to be coordination and exchange of information among all those involved in healthcare. Resistant isolates are seen both in Enterobacteriaceae as well as in gram positive bacteria. The most commonly isolated bacteria are *E.coli* and *Klebsiella*. Among *E.coli* the resistance to commonly used antibiotics is on the rise. A study conducted in 2014 reports a high degree of resistance to Ampicillin (88.4%) followed by Amoxicillin-clavulanic acid (74.4%), Norfloxacin (74.2%), Cefuroxime (72.2%), Ceftriaxone (71.4%) and Cotrimoxazole (64.2%) (Niranjan and Malini, 2014). The antibiotic susceptibility data for uropathogens in India is very minimal. There have been very few community or hospital based studies to assess the problem burden. Our literature search revealed wide variations in the resistance pattern of *E.coli*. The present study was undertaken to assess the antimicrobial susceptibility patterns of *E.coli* in order to guide the clinician regarding presumptive antibiotic usage in UTI patients. There exists lacunae with regard to quantification of the problem and various determining factors related to antimicrobial resistance. There is an urgent need to develop and strengthen antimicrobial policy, standard treatment guidelines, national plan for containment of AMR and research related to public health aspects of AMR at community and hospital level in India (Ganesh Kumar *et al.*, 2013)

Materials and Methods

Freshly void mid-stream urine samples received in the Microbiology laboratory of Azezia Institute of Medical Sciences and Research from January 2017 to December 2017 were included in the study. A total of 1152 urine samples were received during the study period. 327 samples grew monobacterial

significant bacteruria. 611 samples did not show any growth and 173 samples grew more than 3 types of colonies and were hence termed as mixed growth. The samples were initially examined by wet mount. Semi-quantitative cultures were performed using calibrated loop method on 5% Sheep blood agar and Mac Conkey agar. The plates were incubated at 37°C for 18-24 hours. Significance of samples was determined as per Kass criteria (European Association of Urology, 2013). Samples growing single bacterial species with $>10^5$ cfu/ml were considered as significant bacteruria samples. Isolates were identified based on the colony morphology, Gram staining and standard biochemical reactions (Koneman *et al.*, 2006; Gerald Collee *et al.*, 14th edn). The isolates were then subjected to Antibiotic sensitivity testing as per Clinical and Laboratory Standards Institute (CLSI) 2017 guidelines (Performance Standards for Antimicrobial Susceptibility Testing). The Antibiotic sensitivity testing was done on Mueller Hinton agar. Standard strain American Type Culture Collection (ATCC) strain of *E.coli* 25922 was used as a control. The culture media were obtained from Himedia labs, Mumbai, India and were prepared in-house. The prepared plates were approved for use after internal quality control. The antibiotic discs were obtained from Himedia labs as well. The isolates were tested for sensitivity to the following discs Ampicillin (10µg), Gentamicin (10µg), Tobramycin (10µg), Amikacin (30µg), Ciprofloxacin (5µg) Levofloxacin (5µg) Cotrimoxazole (1.25/23.75µg) Amox-Clavulanic Acid (20/10µg) Piperacillin-Tazobactam (100/10µg) Norfloxacin (10µg) Amikacin (30µg) Cefuroxime (30µg), Ceftriaxone (30µg), Ceftazidime (30µg), Ceftazidime-Clavulanic Acid (30/10µg), Cefotaxime (30µg) Imipenam (10µg), Tetracycline (30µg), Nitrofurantoin (300µg). The sensitivity was recorded and reported as per the CLSI 2017

guidelines.

Results and Discussion

Among the 1152 urine samples received for culture and sensitivity 327 samples (28.4%) were identified as significant bacteruria samples. 611 (53.03%) samples did not show any growth after 24hours of incubation. These samples were further incubated for 24 hours and declared as culture negative at the end of 48hours. 173 (15.01%) samples grew more than 3types of colonies after 24hours of incubation. These samples were reported as mixed growth along with comment to repeat sample with Clean Catch Mid-Stream urine (CCMSU). The predominant growth was *E.coli* (148 isolates - 45.29%). 22.67% isolated were identified as Klebsiella pneumoniae. Gram positive bacteruria comprising of Enterococcus sp, Staphylococcus sp and Streptococcus sp constituted 15.90% (52 isolates). The percentage distribution is provided in Table 1. Gender wise, more *E.coli* were isolated from female patients. Among the 148 isolates of *E.coli* in significant bacteruria samples 96 isolates were from female patients and 52 isolates from male patients (Table 2). The proportions of mixed growths were also higher among samples received from female patients. 141 samples out of the total 1152 showed mixed growths in females (12.23%) whereas in males 33 samples were reported as mixed growth (2.86%). Most of the *E.coli* isolates were isolated from in-patients in our study (89 isolates). 31 and 28 isolates were obtained from samples received from out patients and Intensive care units respectively (Table 3). Out of 148 isolates of *E.coli* 96 were obtained from female patients. 69 out of these isolates were from female patients admitted in the hospital. The majority of *E.coli* was isolated from samples received from General medicine department (50.7%). The rest were isolated from samples received from Emergency, Obstetrics and Gynaecology, General Surgery,

Urology, paediatrics and Orthopaedics department (Table 4). In our study the Urinary Tract Infections due to *E.coli* are more common in females from the second decade of life. In males the infection due to *E.coli* was more common after 5th decade (Table 5).

First isolates of each patient were considered for the antibiogram. 129 first time isolate analysis showed sensitivity to Imipenam (91%), Nitrofurantoin (97%), Amikacin (91%), Tobramycin (72%), Piperacillin-Tazobactam (77%), Gentamicin (66%) and Tetracycline (46%). *E.coli* isolated exhibited resistance to Ampicillin, Amoxicillin-Clavulanic acid, 2nd and 3rd generation Cephalosporins and Fluroquinolones. *E.coli* was sensitive to Cotrimoxazole in 49% isolates

The antibiotic sensitivity pattern was assessed for Multi Drug Resistant Organisms (MDRO). Of the 129 first time *E.coli* isolates 50 were sensitive to most drugs (38.7%), 62 isolates

were resistant to at least one antibiotic in three or more groups (48.1%) 17 strains of *E.coli* exhibited resistance to most antibiotics barring two or less groups (13.2%) and were termed as XDR isolates (Koneman *et al.*, 2006). We did not come across any Pan drug resistant strains in this study (Silpi Basak *et al.*, 2016).

Around 150 million UTIs occur yearly on a global basis; resulting in more than 6 billion dollars in direct health care expenditures. Infections are rare in boys except in association with anatomic or functional abnormalities in the first year of life.

Infections are also infrequent among 2- to 13-year-old girls, but some young girls experience multiple repeated episodes of recurrent cystitis or pyelonephritis. The incidence of UTIs markedly increases among young women during adolescence. As per the study carried out by Prinja *et al.*, Rs.1713 is spent per head towards healthcare expenditure annually (Prinja *et al.*, 2012).

Table.1 Distribution of isolates in the urine samples (N=1152)

Isolates	Number (N=366)	Percentage (%)
<i>E.coli</i>	148	40.44
<i>Klebsiella pneumoniae</i>	74	20.22
<i>Enterococcus sp</i>	27	7.38
<i>Staphylococcus aureus</i>	16	4.37
<i>Pseudomonas aeruginosa</i>	15	4.10
<i>Coagulase Negative Staphylococcus aureus</i>	3	0.82
<i>Enterobacter aerogenes</i>	14	3.83
<i>Acinetobacter sp</i>	10	2.73
<i>Streptococcus sp</i>	6	1.64
<i>Proteus sp</i>	8	2.19
<i>Citrobacter sp</i>	6	1.64
<i>Candida albicans</i>	12	3.28
<i>Candida non albicans</i>	27	7.38
Total number of reportable growth	366	100.00
Non reportable growths		
No growth	612	53.1
Mixed growth	174	15.1

Table.2 Gender Distribution of isolates in the urine samples (N=1152)			
Isolates	Number (N=366)	Males	Females
<i>E.coli</i>	148	52	96
<i>Klebsiella pneumoniae</i>	74	36	38
<i>Enterococcus sp</i>	27	12	15
<i>Staphylococcus aureus</i>	16	2	14
<i>Pseudomonas sp</i>	15	11	4
CoNS	3	0	3
<i>Enterobacter sp</i>	14	10	4
<i>Acinetobacter sp</i>	10	6	4
<i>Streptococcus sp</i>	6	2	4
<i>Proteus sp</i>	8	6	2
<i>Citrobacter sp</i>	6	2	4
<i>Candida albicans</i>	12	9	3
<i>Candida non albicans</i>	27	21	6
Total number of reportable growth	366	169	197
Non reportable growth			
No growth	612	218	394
Mixed growth	174	33	141

Table.3 Location wise Distribution of <i>E.coli</i> isolates		
Location type	Female	Male
Inpatients (N=89)	54	35
Intensive care (N=28)	15	13
Outpatients (N=31)	27	4
Total (N=148)	96	52

Table.4 Department wise distribution of <i>E.coli</i> isolates		
Department	Number	Percentage (%)
General medicine	75	50.7
Emergency	16	10.8
Obstetrics and Gynaecology	15	10.1
Urology	13	8.8
General surgery	11	7.4
Orthopaedics	8	5.4
Paediatrics	4	2.7
Neurology	2	1.4
Cardiology	2	1.4
Pulmonology	1	0.7
Nephrology	1	0.7

Table.5 Age-wise distribution of *E.coli* isolates (N=148)

Age category	Number		Percentage	
	Males	Females	Males	Females
<1	1	0	0.68	0.00
1-10years	2	3	1.35	2.03
11-20Years	0	2	0.00	1.35
21-30years	0	14	0.00	9.46
31-40years	0	12	0.00	8.11
41-50years	1	11	0.68	7.43
51-60years	13	14	8.78	9.46
61-70years	11	16	7.43	10.81
71-80years	13	15	8.78	10.14
81-90years	10	9	6.76	6.08
>91years	1	0	0.68	0.00

This is bound to go up with drug resistant *E.coli* causing UTI. *E.coli* is the most common organism isolated from urinary tract infections in India as well (Atul Kothari and Vishal Sagar, 2008). The rate of culture positivity is quite low in most of the studies. In our study 611 samples out of the 1152 samples received did not grow any bacteria. The possible explanation for this can be incorrect timing of the sample collection or sample may have been taken after administration of antibiotic. In the current scenario of over the counter medications most patients tend to start antibiotics even before they report to health care facility. 169 samples were reported as mixed growth. This could be minimized by giving proper instructions are given to the patients regarding sample collection (CCMSU). Secondly in our study most mixed growth samples were seen in females which may be due to anatomical proximity of gastrointestinal tract to genitourinary tract in females. Nevertheless proper instructions on Ano-genital toilet and proper timing of sample collection can bring down the proportion of non-reportable culture growths.

Among the samples showing significant monobacterial growth *E.coli* was

predominant. Age wise split showed that the Urinary tract infections in female patients were more common from 2nd to 5th decade of life. In males the incidence of urinary tract infections due to *E.coli* was more predominant after 5th decade of life (Devanand Prakash and Ramchandra Sahai Saxena, 2013).

In our study *E.coli* showed high degree of resistance to Cephalosporins and Fluroquinolones. This correlates with the prescribing trends prevailing in this part of the country. Now the million dollar question is how to avoid resistance. There have been many suggestions, recommendations from international agencies (WHO and CDC). They recommended strict control on antibiotic use by humans. A prescription should be mandatory for procuring the antibiotic. Secondly the international agencies have also called for controlled therapeutic use of antibiotics in animal husbandry. The next method suggested in antibiotic cycling. For this every healthcare unit needs to develop their own antibiogram to have a clear picture of the antibiotic resistance pattern prevalent in the area. Antibiotic cycling is a short term measure to tackle antibiotic resistance and also to prolong the usefulness of live saving

drugs. This procedure will reduce the selection pressure for resistance (Silpi Basak *et al.*, 2016).

The Chennai declaration documented the fact of emergence of antibiotic resistance in India and it aimed to form a “Road map” to tackle the menace of antibiotic resistance in India using a stepwise approach (Ghafur *et al.*, 2012). The uniform consensus was that there is paucity of published data and antibiotic usage and antibiotic resistance in India. The road map included regulation of over the counter sale of antibiotics, in-hospital antibiotic usage monitoring and stringent supervision on usage of life-saving drugs. It was also decided that ICMR would set up a National antibiotic resistance surveillance system with representations from all parts of the country (Magiorakos *et al.*, 2012). The ICMR has come out with Treatment guidelines for antimicrobial use in common syndromes – 2017. These were based on data obtained from central institutes (PGIMER, AIIMS, JIPMER, and CMC Vellore) and available national data. These guidelines show high degree of resistance to Cefotaxime (72-87%), Ceftazidime (72-89%) and Ciprofloxacin (74-90%). As per the recommendation issued by ICMR Nitrofurantoin, Cotrimoxazole and Ciprofloxacin have been suggested as first line drugs for acute uncomplicated cystitis in the absence of cultures. In our Study *E.coli* were sensitive to Nitrofurantoin. There was high degree of resistance to Cotrimoxazole (55%) and Ciprofloxacin (63%). The effectiveness of these drugs in treatment of cystitis will have to be relooked and assessed based on the national level data from surveillance network.

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